

Apalachicola (APA) NERR Nutrient Metadata
January – December 2016
Latest Update: January 24, 2018

I. Data Set and Research Descriptors

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2) Research objectives –

Previous studies have shown the importance of river flow and flushing rates on nutrients and primary productivity in Apalachicola Bay. Similar studies have determined nitrogen and phosphorus budgets as well as nutrient limitations related to seasonality and river flow (Elder and Mattraw 1982, Frick et al. 1996, Mortazavi 1998, Twilley et al. 1999, Mortazavi 2000a, b, Mortazavi et al. 2001, Putland 2005, Edmiston 2008, Caffrey et al. 2013). There has been an ongoing controversy between the states of Florida, Georgia, and Alabama over the upstream diversion of water for 25 years. Approximately 88% of the Apalachicola River and Bay drainage basin is located in Georgia and Alabama and historical flows are being threatened by upstream use. A tri-state compact between the states and approved by the US Congress, required negotiations between the states to develop a water allocation formula. The states were unable to come to an agreement and the compact expired. In late 2014, the US Supreme Court agreed to hear the case and legal proceedings are currently underway. The research objectives of this study are to investigate short-term variability, long-term change, and the relationship of other environmental factors to the productivity of the Apalachicola Bay system as well as try to separate natural from man-made variability. Data from this monitoring project has also been used by Florida DEP in support of Numeric Nutrient Criteria development.

a) Monthly Grab

Monthly grab samples are collected at 11 sites located across Apalachicola Bay to monitor spatial and temporal fluctuations in nutrient and chlorophyll-*a* concentrations across the bay. The stations were chosen to help determine the influence of the river, local rainfall, adjacent habitats and anthropogenic impacts on the Bay. Sampling sites are located in the lower Apalachicola River, in the coastal area, offshore of the barrier islands, at the SWMP datalogger locations (primary SWMP stations), and throughout the bay. Seasonal, climatic, and anthropogenic factors all impact river flow, which in turn affects nutrient and chlorophyll-*a* concentrations in the bay. Nutrient and chlorophyll-*a* concentrations are also influenced by biological activity, tidal action, wind direction and speed, and the hydrodynamics of the system.

b) Diel Sampling Program

Diel sampling is performed once a month in conjunction with grab sampling for nutrients and chlorophyll-*a* concentration. The East Bay Surface water quality datalogger site (apaesnut) is utilized each month for placement of the sampler so that temporal water quality data may be compared with the spatial nutrient and chlorophyll *a* data collected at this site. Studies by the Reserve and others have shown the influence of tidal action and runoff on other physical parameters in the bay (Estabrook 1973, Livingston 1978, Livingston and Duncan 1979, Edmiston 2008). Diel samples are collected over a 25-hour period thereby covering the lunar day of 24 hours 48 minutes.

3) Research methods –

a) Monthly Grab Sampling Program

Monthly grab samples are collected at eleven stations (see Table 1) within and adjacent to Apalachicola Bay. All grab samples are collected on the same day. Because of the distance between the stations it is not always possible to collect all the samples several hours prior to low tide. Tidal condition, wave height, wind direction, speed, precipitation, and cloud cover are recorded for each station at the time of sampling but are not included in this dataset and are available upon request. Climatic data from the Apalachicola National Estuarine Research Reserve (ANERR) weather station is available online at www.nerrsdata.org. Sampling after heavy rains is avoided if possible. Water temperature, salinity, specific conductivity, dissolved oxygen, pH, total dissolved solids, and turbidity are measured at surface and bottom for each station with a YSI Pro DSS handheld meter. Surface measurements only are included in this dataset for temperature, salinity, pH, and dissolved oxygen (with the exception of the East Bay Bottom station). Bottom measurements for temperature, salinity, specific conductivity, dissolved oxygen, pH, total dissolved solids, and turbidity are available on request. Secchi data is also included in this dataset. In addition to readings taken by the hand-held instrument, turbidity samples are collected at each site and are analyzed in the ANERR lab with a HR Scientific DRT-15CE Turbidimeter. In 2016, biochemical oxygen demand was measured from whole water samples for the months of March, June, September, and December (quarterly) at all stations except for apaebnut. These data are not included in the dataset but are available

by contacting the Reserve directly. All grab samples are analyzed at the Florida Department of Environmental Protection laboratory (FLDEP).

i) Grab sample collection:

A submersible pump and flexible clear plastic tubing is used to collect water from a depth of 0.5 meters at all stations not associated with a SWMP datalogger site. At the Cat Point and Dry Bar SWMP datalogger stations, water samples are collected at a depth of approximately 1.5 meters below the surface to match the approximate depth of the probes of the data loggers deployed at these sites. At the East Bay datalogger station water samples are collected from surface (0.5 meters) and bottom (1.5 meters) depths, approximating the depths of the two dataloggers deployed at this site. Triplicate samples are collected every other month at one randomly selected SWMP station.

ii) Grab sample filtration and handling:

Water from the submersible pump is delivered directly into the appropriate sample bottles. For samples requiring filtration, an in-line filter is attached to the end of the flexible tubing, and water filtered in this manner is delivered directly to the appropriate sample bottles. Necessary preservatives are added prior to water sample according to appropriate EPA protocols for nutrient sampling. Whole water samples for chlorophyll-*a* analysis are filtered at the FLDEP laboratory. All samples are placed on ice in the dark until delivery to the FLDEP laboratory. The submersible pump and tubing are flushed with ambient water prior to sample collection at each station. If an additional filter is needed at a site, either a new filter holder and filter will be used or the current filter holder is rinsed with DI prior to addition of a new filter. A field blank is also run each month, using deionized water (DI) water for sample blank. The field blank is delivered using the pump, tubing and filter as described above. All grab samples are delivered to the FLDEP laboratory 24 to 36 hours after collection.

b) Diel Sampling Program

Diel sampling is performed with an ISCO 3700 Portable Automated Sampler at the East Bay surface (apaesnut) station. The ISCO is deployed on a fixed platform located at the East Bay surface site. Generally, the ISCO is deployed at the beginning of the grab sample collection trip and retrieved the following morning. In some months, adverse weather conditions result in deployment of the ISCO sampler during a week other than the week of grab sample collection. The sampler is programmed to collect two 1000 ml water samples every 2.5 hours, over a 25-hour period at the same depth as the East Bay surface datalogger probes (0.5 m below surface). This captures a complete 24 hour 48-minute lunar-tidal cycle. The ISCO sampler is programmed to purge the suction line before and after each sample collection. The center of the ISCO sampler is filled with ice to aid in sample preservation. All samples are placed on ice upon retrieval of the ISCO sampler at the end of the sampling period. Nutrient sample filtration is performed at ANERR laboratory within one hour of retrieval from the ISCO sampler. Whole water samples for chlorophyll-*a* analysis are filtered at the FLDEP laboratory. All diel samples are delivered to the FLDEP laboratory within 36 hours of the first sample collection time. Note: No duplicate diel samples are taken, however there is some overlap with monthly grabs collected at the East Bay Surface station and deployment of the ISCO sampler.

c) Equipment QAQC and maintenance – Grab and Diel Sampling Program:

The submersible pump, tubing, and filter holders used in the field are acid rinsed with 10% Hydrochloric Acid and triple rinsed with ultra-pure DI water after each sampling trip. Laboratory items such as the filtration funnels and receivers are acid washed with 10% Hydrochloric Acid and triple rinsed with ultra-pure DI water after each sampling event. Diel sample collection bottles used in the ISCO automated sampler are acid washed and triple rinsed with ultra-pure DI water after each sampling event. The ISCO automated sampler tubing is acid washed and triple rinsed with ultra-pure DI water after each sampling event. The overall condition of the pump and tubing is checked each month prior to deployment and tubing is replaced as needed, and per the CDMO SOP replacement schedule. New, unused sample bottles are supplied by FLDEP laboratory for each grab sampling event. The YSI Pro DSS and Turbidimeter are calibrated before each sampling event.

4) Site location and character –

The Apalachicola Drainage Basin encompasses over 50,700 square kilometers and includes parts of three states (Alabama, Georgia, and Florida). The Apalachicola River is the largest in Florida in terms of flow. The amount of river discharge has

been shown to be highly significant to the ecology of the estuary, which acts as a buffer between the Gulf of Mexico and fresh water input from upland areas. The nutrient rich plume of "green water" moving out of Apalachicola Bay is also important to the productivity of the northeastern Gulf of Mexico. ANERR is located in the northwestern part of Florida, generally called the panhandle. It is located adjacent to the Cities of Apalachicola and Eastpoint, and encompasses most of the Apalachicola Bay system, including 84 kilometers of the lower Apalachicola River. Passes, both natural and manmade, connect Apalachicola Bay to the northeastern Gulf of Mexico. Nutrient discharge and pollutant runoff surrounding the city of Apalachicola is elevated, compared to minimal pollution draining to Apalachicola Bay from the undeveloped panhandle.

Monthly grab samples are collected at all SWMP and nutrient monitoring stations. A map of station locations is given in Figure 1.

a) East Bay datalogger and nutrient station

East Bay is separated from Apalachicola Bay by two bridges and a causeway and is located to the north of the bay proper. East Bay is 8.2 km long, has an average depth of approximately 1.0 m MHW, and an average width of 1.8 km. The tides in East Bay are mixed and range from 0.3 m to 1.0 m (average 0.5 m). The datalogger and nutrient sampling site is located in the upper reaches of East Bay. The tower location for the two East Bay dataloggers (ES and EB) is 29.7858 N, 84.8752 W. At the sampling site, the depth is 2.2 m MHW and the width of the bay is 1.0 km. The tides in the system are mixed, meaning the number of tides can range from one to five tides during a 24-hour period and are not evenly distributed throughout the day. At the East Bay bottom site the meter probes are 1.5 meters below the surface (or 0.3 m off the bottom sediment). Salinity ranges from 0 to 30 psu and the long-term (1995 – 2016) average salinity is 11.2 psu. At the East Bay surface site the meter probes are 0.5 meters below the surface (or 1.7 m off the bottom sediment) and salinity ranges from 0 psu to 30 psu with a long term (1995 – 2016) average salinity of 9.9 psu. The freshwater input is very tannic and usually dark colored. Flows vary with local rainfall and are not quantified due to the diverse sources of the runoff. The bottom habitat at this bay site is soft sediment, primarily silt and clay, with no vegetation present. The dominant marsh vegetation near the sampling site (approximately 300 meters away) is needlerush grass (*Juncus roemerianus*) and swamp sawgrass (*Cladium jamaicense*) and smooth cordgrass (*Spartina alterniflora*). The dominant upland vegetation is primarily pineland forests which includes slash pine (*Pinus elliotii*), saw palmetto (*Serenoa repens*), and sand pine (*Pinus clausa*). Upland land use near the sampling site includes conservation and silviculture uses with some single family residential in the lower East Bay area. The sampling site is influenced by local runoff from Tate's Hell Swamp, the East Bay marshes, and distributary flow, some of which comes from the Apalachicola River via the East River. Tate's Hell Swamp was ditched, diked, and altered in the late 1960's and early 1970's by timber companies. These changes shortened the drainage period and allowed increased runoff with a concomitant decrease in pH and increase in color, which had a drastic effect on the biological communities in East Bay. Restoration of Tate's Hell Swamp began in 1995 to reduce non-point source runoff and restore historic sheet flow in the area.

b) Cat Point datalogger and nutrient station

The Cat Point datalogger and nutrient sampling site is located in St. George Sound, approximately 400 meters east of the St. George Island Bridge. The piling location is 29.7021 N, 84.8802 W. The tides at Cat Point are mixed and range from 0.3 m to 1.0 m (average 0.5 m). At the sampling site, the depth is 2.5 meters MHW, and the width of the bay is 6.4 km. At the Cat Point site the datalogger probes are located 1.5 meters below the surface (or 0.3 m off the bottom sediment). This is also the approximate depth where nutrients are collected monthly. Salinity ranges from 0 to 34 psu with a long-term (2002 – 2016) average salinity of 22.0 psu. Flows vary with local rainfall and are not quantified due to the diverse sources of the runoff. The bottom type is oyster bar with no vegetation present except algae growing on the oysters in the summer. The dominant upland vegetation is primarily pineland forests, which include slash pine (*Pinus elliotii*), saw palmetto (*Serenoa repens*), and sand pine (*Pinus clausa*). Upland land use near the sampling site includes single family residential and commercial use in the Eastpoint area. The sampling site is influenced by local runoff from Tate's Hell Swamp and flow from the Apalachicola River. High salinity water comes mainly from the east, through East Pass at the eastern end of St. George Island.

c) Dry Bar datalogger and nutrient station

The Dry Bar datalogger and nutrient sampling site is located near St. Vincent Sound, in the western part of the Apalachicola Bay system, approximately 0.8 kilometer east of St. Vincent Island. The tower location is 29.6747 N, 85.0584 W. The tides are mixed and range from 0.3 to 1.0 meters. At the sampling site, the depth is 2.0 meters MHW and the width of the bay is 11.2 km. At the Dry Bar site the datalogger probes are located 1.5 meters below the surface (or 0.3 m off the bottom sediment). This is also the approximate depth where nutrients are collected monthly. Salinity ranges from 0 to 34 psu with a long-term (2002 – 2016) average salinity of 21.8 psu. Flows vary with local rainfall and are not quantified because the sampling site is influenced by the flow of the Apalachicola River and high salinity water coming through West Pass and Sikes Cut. The bottom type is oyster bar with no vegetation present, except algae that grows on the oysters during the summer months. The dominant upland vegetation includes slash pine (*Pinus clausa*) flatwoods with various combinations of gallberry (*Ilex glabra*), smooth cordgrass (*Spartina alterniflora*), fetterbush (*Leucothoe racemosa*), cabbage palm (*Sabal palmetto*), saw palmetto (*Serenoa repens*), magnolia (*Magnolia grandiflora*), and grasses. Upland use near the sampling site includes state owned and managed Cape St. George Island, St. Vincent National Wildlife Refuge, as well as single family residential and commercial use in the Apalachicola area.

d) Secondary SWMP stations

Detailed information for an additional 7 nutrient (secondary SWMP) stations, not associated with the required sampling at the primary SWMP datalogger sites, as well as the datalogger sites, is included in Table 1.

West Pass

29.6379 N, 85.0890 W

Salinity average = 22.5 psu, range = 1.8 – 36.0 psu

This site is located in the pass between two uninhabited barrier islands, the state owned and managed Cape St. George Island and St. Vincent National Wildlife Refuge. The sampling site is influenced by the flow of the Apalachicola River and high salinity water coming through West Pass.

Pilots Cove

29.6079 N, 85.0196 W

Salinity average = 22.9 psu, range = 1.3 – 35.5 psu

This site is located near state owned and managed Cape St. George Island, an uninhabited barrier island. The sampling site is influenced by the flow of the Apalachicola River and high salinity water coming through West Pass.

Mid Bay

29.6677 N, 84.9940 W

Salinity average = 16.3 psu, range = 0.2 – 35.2 psu

This sampling site is located in central Apalachicola Bay. The site is roughly equidistant from state owned and managed Cape St. George Island (four miles distant), St. Vincent National Wildlife Refuge (six miles distant), and single family residential and commercial use in the Apalachicola area (four miles distant). This site is approximately 2.5 kilometers from the intercoastal waterway channel. The sampling site is influenced by the flow of the Apalachicola River and high salinity water coming through Sikes Cut and West Pass.

East Bay Bridge

29.7308 N, 84.9452 W

Salinity average = 7.9 psu, range = 0 – 30.7 psu

This site is located near the western section of the US Highway 98 bridge connecting Apalachicola and Eastpoint. The bridge also serves as the boundary line between East Bay and Apalachicola Bay. Nearby upland areas consist of residential and commercial use in the areas surrounding the cities of Apalachicola and Eastpoint. The sampling site is influenced by flows from the Apalachicola River and tributaries including the Little St. Marks River, St. Marks River, and East River.

Sikes Cut offshore

29.6067 N, 84.9467 W

Salinity average = 31.9 psu, range 21.7 – 35.8 psu

This site is selected to characterize true marine water, and is located south of Sikes Cut in the Gulf of Mexico. The site is near the eastern portion of state owned and managed Cape St. George Island and near the western end of St. George Island in an area consisting of single family and vacation homes. Sikes Cut allows tidal exchange of high salinity water from the Gulf of Mexico and lower salinity water from Apalachicola Bay. Sikes Cut is an important pass utilized by commercial and recreational vessels.

Nicks Hole

29.6504 N, 84.9289 W

Salinity average = 19.0 psu, range = 0.5 – 35.4 psu

This site is near single family and vacation home use on St George Island. A small airport utilized by private aircraft is also located near Nicks Hole. The site is tidally influenced by high salinity water from Sikes Cut and by flows from the Apalachicola River.

River

29.7791 N, 85.0434 W

Salinity average = 0.1 psu, range = 0 – 0.1 psu

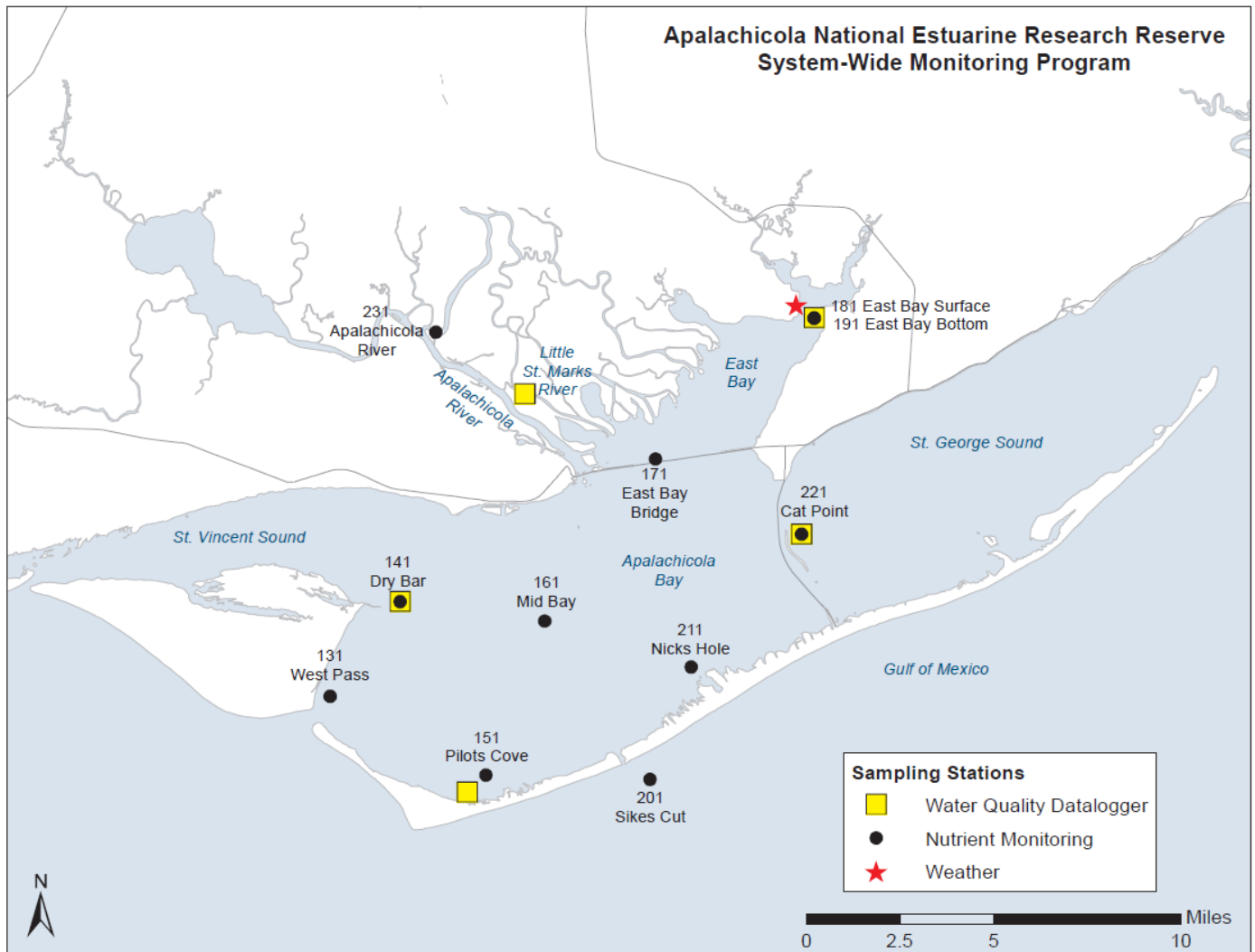
This site is selected to characterize fresh water in the Apalachicola River. The site is located in the central channel of the river approximately 9.5 kilometers north and upstream of the river mouth and the residential and commercial areas of Apalachicola. Adjacent areas are state owned and managed forested floodplain. The site is influenced by Apalachicola River flow.

Table 1. Nutrient and chlorophyll *a* sampling sites for the Apalachicola NERR SWMP.

Station code	SWMP Status	Station name	Location	Active Dates	Tidal Range Average (meters)	Salinity Range	Water Depth Average (meters)	Bottom Habitat	Datalogger Station Name	Sample Depth (meters)	Reason Decommissioned	Notes
apawpnut	S	West Pass	29° 38' 16.44 N, 85° 5' 20.40 W	04/01/2002 - current	0.7	euryhaline	5.0	sand		0.5	NA	NA
apadbnut	P	Dry Bar	29° 40' 28.92 N, 85° 3' 29.88 W	04/01/2002 - current	0.7	euryhaline	1.7	oyster bar	apadbwq	1.5	NA	NA
apapcnut	S	Pilot's Cove	29° 36' 28.44 N, 85° 1' 10.56 W	04/01/2002 - current	0.7	euryhaline	1.8	patchy seagrass		0.5	NA	NA
apambnut	S	Mid Bay	29° 40' 3.72 N, 84° 59' 38.40 W	04/01/2002 - current	0.7	euryhaline	2.2	sandy silt		0.5	NA	NA
apaegnut	S	East Bay Bridge	29° 43' 50.88 N, 84° 56' 42.72 W	04/01/2002 - current	0.7	euryhaline	1.6	silty clay		0.5	NA	NA
apaesnut	P	East Bay Surface	29° 47' 8.88 N, 84° 52' 30.72 W	04/01/2002 - current	0.7	euryhaline	1.7	clayey sand	apaeswq	0.5	NA	NA
apaebnut	P	East Bay Bottom	29° 47' 8.88 N, 84° 52' 30.72 W	04/01/2002 - current	0.7	euryhaline	1.7	clayey sand	apaebwq	1.5	NA	NA
apascnut	S	Sikes Cut Offshore	29° 36' 24.12 N, 84° 56' 48.12 W	04/01/2002 - current	0.7	marine	>5.0	sand		0.5	NA	NA
apanhnut	S	Nick's Hole	29° 39' 1.44 N, 84° 55' 44.04 W	04/01/2002 - current	0.7	euryhaline	1.0	patchy seagrass		0.5	NA	NA
apacpnut	P	Cat Point	29° 42' 7.68 N, 84° 52' 48.72 W	04/01/2002 - current	0.7	euryhaline	1.8	oyster bar	apacpwq	1.5	NA	NA
aparvnut	S	River	29° 46' 44.76 N, 85° 2' 36.24 W	04/01/2002 - current	0.7	oligohaline	3-4	sandy silt		0.5	NA	NA

Note: Diel samples are collected 2.5 hours apart at the East Bay Surface datalogger site, APAESNUT, with the ISCO automated water sampler. No duplicate diel samples are taken, however there is some overlap with monthly grabs collected at the East Bay Surface station and deployment of the ISCO sampler.

Figure 1: ANERR SWMP Station locations.



4) Coded variable definitions –

Station code names:

apacpnut = Apalachicola Reserve nutrient data for Cat Point
 apadbnut = Apalachicola Reserve nutrient data for Dry Bar
 apaebnut = Apalachicola Reserve nutrient data for East Bay Bottom
 apaegnnt = Apalachicola Reserve nutrient data for East Bay Bridge
 apaesnnt = Apalachicola Reserve nutrient data for East Bay Surface
 apambnut = Apalachicola Reserve nutrient data for Mid Bay
 apahnnt = Apalachicola Reserve nutrient data for Nicks Hole
 apapcnut = Apalachicola Reserve nutrient data for Pilots Cove
 aparvnnt = Apalachicola Reserve nutrient data for River

apascnut = Apalachicola Reserve nutrient data for Sikes Cut
 apawpnut = Apalachicola Reserve nutrient data for West Pass

Monitoring Programs:
 Monthly grab samples = 1
 Diel grab sampling = 2

6) Data collection period –

Nutrient monitoring began in April 2002 at all stations listed. Sampling has been performed monthly at all stations, unless otherwise noted. This table lists collection times for all nutrient and chlorophyll-*a* samples in 2016. The Start and End date and times listed below reflect the times that the first and last diel samples were collected for each monthly diel sampling event. Grab sample end time is not recorded in the field. Grab sample collection, filtering, and icing are completed within 10 minutes or less depending upon field conditions at the time of sampling. Time is coded based on a 2400 hour clock and is referenced to Eastern Standard Time (EST), without Daylight Savings Time adjustments.

a) Samples date/times Monitoring Program 1 (Grab Samples)

Site	Date	Time	Site	Date	Time	Site	Date	Time
apacpnut	01/13/16	12:13	apadbnut	01/13/16	9:10	apaebnut	01/13/16	9:05
apacpnut	02/17/16	11:18	apadbnut	02/17/16	8:48	apaebnut	02/17/16	11:57
apacpnut	03/01/16	9:59	apadbnut	03/01/16	13:25	apaebnut	03/01/16	8:31
apacpnut	04/05/16	9:37	apadbnut	03/01/16	13:26	apaebnut	04/05/16	8:10
apacpnut	05/10/16	8:48	apadbnut	03/01/16	13:27	apaebnut	05/10/16	7:56
apacpnut	05/10/16	8:49	apadbnut	04/05/16	12:22	apaebnut	06/01/16	7:36
apacpnut	05/10/16	8:50	apadbnut	05/10/16	10:30	apaebnut	07/05/16	7:27
apacpnut	06/01/16	8:44	apadbnut	06/01/16	11:28	apaebnut	07/05/16	7:28
apacpnut	07/05/16	8:32	apadbnut	07/05/16	10:15	apaebnut	07/05/16	7:29
apacpnut	8/2/16	8:29	apadbnut	8/2/16	10:47	apaebnut	8/2/16	7:38
apacpnut	9/12/16	9:05	apadbnut	9/13/16	7:16	apaebnut	9/12/16	7:52
apacpnut	9/12/16	9:07	apadbnut	10/3/16	9:42	apaebnut	10/3/16	7:40
apacpnut	9/12/16	9:09	apadbnut	11/1/16	10:19	apaebnut	11/1/16	7:50
apacpnut	10/3/16	8:14	apadbnut	12/7/16	13:52	apaebnut	12/7/16	8:44
apacpnut	11/1/16	8:34						
apacpnut	12/7/16	10:22						
Site	Date	Time	Site	Date	Time	Site	Date	Time
apaegnut	01/13/16	12:43	apaesnut	01/13/16	9:08	apambnut	01/13/16	8:48
apaegnut	02/17/16	12:36	apaesnut	02/17/16	11:57	apambnut	02/17/16	8:30
apaegnut	03/01/16	9:24	apaesnut	03/01/16	8:31	apambnut	03/01/16	14:00
apaegnut	04/05/16	9:05	apaesnut	04/05/16	8:10	apambnut	04/05/16	13:11
apaegnut	05/10/16	8:27	apaesnut	05/10/16	7:55	apambnut	05/10/16	11:00
apaegnut	06/01/16	8:15	apaesnut	06/01/16	7:35	apambnut	06/01/16	12:06
apaegnut	07/05/16	8:05	apaesnut	07/05/16	7:26	apambnut	07/05/16	10:36
apaegnut	8/2/16	8:10	apaesnut	8/2/16	7:38	apambnut	8/2/16	11:08
apaegnut	9/12/16	8:30	apaesnut	9/12/16	7:50	apambnut	9/13/16	8:16

apaegnnt	10/3/16	7:58	apaesnut	10/3/16	7:38	apambnut	10/3/16	10:00
apaegnnt	11/1/16	8:15	apaesnut	11/1/16	7:48	apambnut	11/1/16	10:41
apaegnnt	12/7/16	9:42	apaesnut	11/1/16	7:52	apambnut	12/7/16	14:23
			apaesnut	11/1/16	7:54			
			apaesnut	12/7/16	8:44			
Site	Date	Time	Site	Date	Time	Site	Date	Time
apanhnut	01/13/16	11:41	apapcnut	01/13/16	10:48	aparvnut	01/13/16	13:18
apanhnut	01/13/16	11:44	apapcnut	02/17/16	10:13	aparvnut	02/17/16	13:06
apanhnut	01/13/16	11:48	apapcnut	03/01/16	12:12	aparvnut	03/01/16	14:41
apanhnut	02/17/16	10:59	apapcnut	04/05/16	11:01	aparvnut	04/05/16	13:10
apanhnut	03/01/16	10:34	apapcnut	05/10/16	9:48	aparvnut	05/10/16	11:45
apanhnut	04/05/16	10:10	apapcnut	06/01/16	10:25	aparvnut	06/01/16	12:46
apanhnut	05/10/16	9:14	apapcnut	07/05/16	9:37	aparvnut	07/05/16	11:05
apanhnut	06/01/16	9:17	apapcnut	8/2/16	9:50	aparvnut	8/2/16	12:06
apanhnut	07/05/16	8:50	apapcnut	9/12/16	10:46	aparvnut	9/13/16	9:09
apanhnut	8/2/16	8:52	apapcnut	10/3/16	9:06	aparvnut	10/3/16	10:30
apanhnut	9/12/16	9:51	apapcnut	11/1/16	9:34	aparvnut	11/1/16	11:10
apanhnut	10/3/16	8:28	apapcnut	12/7/16	12:14	aparvnut	12/7/16	15:15
apanhnut	11/1/16	8:57						
apanhnut	12/7/16	10:56						

Site	Date	Time	Site	Date	Time
apascnut	01/13/16	11:16	apawpnut	01/13/16	10:07
apascnut	02/17/16	10:36	apawpnut	02/17/16	9:40
apascnut	03/01/16	11:27	apawpnut	03/01/16	12:45
apascnut	04/05/16	10:42	apawpnut	04/05/16	12:03
apascnut	05/10/16	9:02*	apawpnut	05/10/16	10:06
apascnut	06/01/16	9:56	apawpnut	06/01/16	10:56
apascnut	07/05/16	9:15	apawpnut	07/05/16	9:56
apascnut	8/2/16	9:25	apawpnut	8/2/2016	10:24
apascnut	9/12/16	10:10*	apawpnut	09/13/16	11:15*
apascnut	10/3/16	8:49	apawpnut	10/3/16	9:21
apascnut	11/1/16	11:15*	apawpnut	11/1/16	9:52
apascnut	12/7/16	11:43	apawpnut	12/7/16	13:10

*samples marked with an * were not collected due to poor weather conditions.

b) Start and End Date/Time for Monitoring Program 2 (Diel Sampling)

Site	Start Date	Start Time	End Date	End Time
apaesnut	01/13/16	9:00	01/14/16	10:00
apaesnut	02/16/16	9:05	02/17/16	10:05

apaesnut	03/01/16	8:35	03/02/16	09:35
apaesnut	04/05/16	8:15	04/06/16	9:15
apaesnut	05/10/16	8:15	05/11/16	9:15
apaesnut	06/01/16	7:40	06/02/16	8:40
apaesnut	07/05/16	8:40	07/06/16	9:40
apaesnut	08/02/16	7:45	08/03/16	8:45
apaesnut	09/12/16	8:00	09/13/16	9:00
apaesnut	10/03/16	7:45	10/04/16	8:45
apaesnut	11/07/16	10:45	11/08/16	11:45
apaesnut	12/07/16	09:00	12/08/16	10:00

7) Associated researchers and projects–

The Reserve conducts long-term water quality monitoring and maintains a weather station as part of the NERRS SWMP. SWMP water quality and weather data may be accessed via www.nerrsdata.org. Other ongoing projects or data that relate to the nutrient monitoring project include:

Apalachicola Bay Oyster Situation Report TP200. UF/IFAS, Sea Grant Florida. April 24, 2013.

Apalachicola River Discharge, U.S. Geological Survey, <http://waterdata.usgs.gov/nwis/>. Ongoing.

Bourque, E., Domangue, R., Lamb, M., Harper, J., Levi, L., Garwood, J., Garland, H., Apalachicola National Estuarine Research Reserve, System Wide Monitoring Program, Long-Term Water Quality Monitoring. Ongoing.

Caffrey, J. University of West Florida. Effect of diurnal and weekly water column hypoxic events on nitrification and nitrogen transformations in estuarine sediments. 2008.

Cannonier, S. Florida Agricultural and Mechanical University School of the Environment, Doctoral Dissertation, HAB Biotxin Concentration in two NERR sites in correlation to nutrient concentrations. Ongoing.

Garwood, J., Domangue, R., Lamb, M., Levi, L., Bourque, E., Garland, H., Cox, N., Christopher, M., Yuan, S., Apalachicola National Estuarine Research Reserve, Distribution and density of fishes and benthic invertebrates in Apalachicola Bay. Ongoing.

Garwood, J., Domangue, R., Lamb, M., Levi, L., Bourque, E., Garland, H., Apalachicola National Estuarine Research Reserve, Effects of River Flow on Estuarine Primary Productivity and Macrozooplankton Communities. Ongoing.

Garwood, J., Levi, L., Domangue, R., Apalachicola National Estuarine Research Reserve, System Wide Monitoring Program, Long-Term Meteorological Monitoring. Ongoing.

Geyer, N. Florida State University, Doctoral Dissertation, Spatio-temporal dynamics of phytoplankton distribution in Apalachicola Bay. 2017.

Harper, J., Domangue, R., Wren, K., Jones, D., Garwood, J., Canedo, J., Snyder, C., Levi, L., Garland, H., NERRS Sentinel Sites Program for Understanding Climate Change Impacts on Estuaries. Ongoing.

Hagen, S., DeLorme, D., Walters, L., Wang, D., Weishampel, J., Yeh, G., Huang, W., Slinn, D., Morris, J. Predicting impacts of sea level rise in the northern Gulf of Mexico. 2015.

Kimbro, D., Garland, H., Christopher, M., Cox, N., Yuan, S., Peter, K., Lamb, M., Harper, J. Apalachicola National Estuarine Research Reserve, Oyster reef research in Apalachicola Bay provided for the ACF lawsuit. 2013-2016.

Site-Specific Information in Support of Establishing Numeric Nutrient Criteria in Apalachicola Bay, Nutrient Criteria Technical Support Document. Division of Assessment and Restoration, Florida Department of Environmental Protection, July 2013.

Tucker, K., Florida Agricultural and Mechanical University Department of Civil and Environmental Engineering, Master's Thesis, Effects of river flow and rainfall on chlorophyll a in Apalachicola River. 2011.

Tucker, K., Florida Agricultural and Mechanical University Department of Civil and Environmental Engineering, Doctoral Dissertation, Nutrient input effects on *Karenia brevis* and *Pseudo-nitzschia* and subsequent marine mortalities in the Gulf of Mexico, Ongoing.

Viveros, P., NOAA Graduate Research Fellowship, University of Florida, Phytoplankton composition and abundance in relation to salinity, nutrient and light gradients in the Apalachicola National Estuarine Research Reserve. 2011.

Wang, H., W. Huang, M. Harwell, L. Edmiston, E. Johnson, P. Hsieh, K. Milla, J. Christensen, J. Stewart, X. Liu. 2008. Modeling oyster growth rate by coupling oyster population and hydrodynamic models for Apalachicola Bay, Florida, USA. Ecological Modeling 211:77-89.

8) Distribution –

NOAA retains the right to analyze, synthesize and publish summaries of the NERRS System-wide Monitoring Program data. The NERRS retains the right to be fully credited for having collected and process the data. Following academic courtesy standards, the NERR site where the data were collected should be contacted and fully acknowledged in any subsequent publications in which any part of the data are used. The data set enclosed within this package/transmission is only as good as the quality assurance and quality control procedures outlined by the enclosed metadata reporting statement. The user bears all responsibility for its subsequent use/misuse in any further analyses or comparisons. The Federal government does not assume liability to the Recipient or third persons, nor will the Federal government reimburse or indemnify the Recipient for its liability due to any losses resulting in any way from the use of this data.

Requested citation format:

NOAA National Estuarine Research Reserve System (NERRS). System-wide Monitoring Program. Data accessed from the NOAA NERRS Centralized Data Management Office website: www.nerrsdata.org; accessed 12 October 2016.

NERR nutrient data and metadata can be obtained from the Research Coordinator at the individual NERR site (please see Principal investigators and contact persons), from the Data Manager at the Centralized Data Management Office (please see personnel directory under the general information link on the CDMO home page) and online at the CDMO home page www.nerrsdata.org. Data are available in comma separated version format.

II. Physical Structure Descriptors

9) Entry verification –

Nutrient data are entered into a Microsoft Excel worksheet and processed using the NutrientQAQC Excel macro. The NutrientQAQC macro sets up the data worksheet, metadata worksheets, and MDL worksheet; adds chosen parameters and facilitates data entry; allows the user to set the number of significant figures to be reported for each parameter and rounds using banker's rounding rules; allows the user to input MDL values and then automatically

flags/codes measured values below MDL and inserts the MDL; calculates parameters chosen by the user and automatically flags/codes for component values below MDL, negative calculated values, and missing data; allows the user to apply QAQC flags and codes to the data; produces summary statistics; graphs selected parameters for review; and exports the resulting data file to the CDMO for tertiary QAQC and assimilation into the CDMO's authoritative online database.

From January – August 2016, Hanna Garland and Lauren Levi were responsible for these tasks. From September – December 2016, Megan Lamb was responsible for these tasks. Megan is the main contact at ANERR for the nutrient program.

10) Parameter titles and variable names by category –

Required NOAA/NERRS System-wide Monitoring Program nutrient parameters are denoted by an asterisk “*”.

Data Category	Parameter	Variable Name	Units of Measure	Collection Period
Phosphorus:				
	*Orthophosphate, filtered	PO4F	mg/L as P	Jan-Dec 2016
	Total Phosphorus	TP	mg/L as P	Jan-Dec 2016
Nitrogen:				
	*Nitrite + Nitrate, filtered	NO23F	mg/L as N	Jan-Dec 2016
	*Ammonium, filtered	NH4F	mg/L as N	Jan-Dec 2016
	Dissolved Inorganic Nitrogen	DIN	mg/L as N	Jan-Dec 2016
	Total Kjeldahl Nitrogen whole	TKN	mg/L as N	Jan-Dec 2016
	Total Nitrogen	TN	mg/L as N	Jan-Dec 2016
Plant Pigments:				
	*Chlorophyll- <i>a</i>	CHLA_N	µg/ L	Jan-Dec 2016
	Uncorrected Chlorophyll- <i>a</i>	UncCHLA_N	µg/ L	Jan-Dec 2016
	Phaeophytin	PHEA	µg/ L	Jan-Dec 2016
Other Laboratory Parameters:				
	Total Suspended Solids	TSS	mg/L	Jan-Dec 2016
Field Parameters:				
	Water temperature	WTEM_N	°C	Jan-Dec 2016
	Salinity	SALT_N	ppt	Jan-Dec 2016
	Dissolved oxygen	DO_N	mg/L	Jan-Dec 2016
	% Saturated dissolved oxygen	DO_S_N	%	Jan-Dec 2016
	pH	PH_N	SU	Jan-Dec 2016
	Turbidity	TURB_N	NTU	Jan-Dec 2016
	Secchi Disk Depth	SECCHI	meters	Jan-Dec 2016

Notes:

1. Time is coded based on a 2400 hour clock and is referenced to Standard Time.
2. Reserves have the option of measuring either NO₂ and NO₃ or they may substitute NO₂₃ for individual analyses if they can show that NO₂ is a minor component relative to NO₃. ANERR has shown NO₂ to be a minor component of NO₂₃.

11) Measured or calculated laboratory parameters –

a) Parameters Measured Directly

Nitrogen species:	NO23F, NH4F, TKN
Phosphorus species:	PO4F, TP
Other:	UncCHLA_N, CHLA_N, PHEA, TSS

b) Calculated Parameters

DIN:	NO23F + NH4F
TN:	NO23F + TKN

12) Limits of detection –

All information in this section is provided by FLDEP laboratory.

a) FLDEP laboratory MDL determination:

MDLs are set such that the risk of reporting a false positive is less than 1%. MDLs are determined using the method specified in the Federal Register, 40 CFR Part 136 Appendix B, using LCSs prepared near the estimated detection limit as surrogates to estimate methodological noise for censored methods (e.g., chromatographic methods which censor analytical noise) or, for uncensored methods, using actual method blanks to directly measure methodological noise. Where the possibility exists for significant systematic bias from sample preparation and handling or from the analytical determinative step (typically inorganic analyses), bias is taken into account when calculating detection limits. Published MDLs may be set higher than experimentally determined MDLs to (1) avoid observed positive interferences from matrix effects or common reagent contaminants or (2) for reporting convenience (i.e., to group common compounds with similar but slightly different experimentally determined MDLs). MDLs are determined in a suitable analyte-free matrix when possible. For certain analytes and matrices, no suitable, analyte-free matrix may be available. In those cases, MDLs are determined in the absence of any matrix, but in the presence of all preparatory reagents carried through the full preparatory and determinative steps. LOD verification procedures may be found in SOP LB-031, Limit of Detection Verification. (From page 39 of FLDEP Laboratory Quality Manual 2016 located at: http://publicfiles.dep.state.fl.us/dear/labs/lab_qualitymanual_16.pdf)

b) 2016 MDLs for Orthophosphate (PO4F), Total Phosphorus (TP), Nitrate + Nitrite (NO23F), ammonium (NH4F), Total Kjeldahl Nitrogen whole (TKN), and Total Suspended Solids (TSS), as reported by FLDEP laboratory:

Parameter	Start Date	End Date	MDL	Units	Date Revisited
PO4F	01/01/16	12/31/16	0.0040	mg/L as P	6/6/2015
TP	01/01/16	12/31/16	0.0040	mg/L as P	6/6/2015
NO23F	01/01/16	12/31/16	0.0040	mg/L as N	6/6/2015
NH4F	01/01/16	11/30/16	0.0020	mg/L as N	6/6/2015
NH4F	12/01/16	12/31/16	0.0020	mg/L as N	6/6/2015
TKN	01/01/16	12/31/16	0.16	mg/L as N	6/6/2015
TSS	01/01/16	01/31/16	2.0	mg/L	6/6/2015
TSS	02/01/16	12/31/16	3.0	mg/L	6/6/2015

FLDEP MDLs for the chlorophyll suite of components may change by station and month based on the need to dilute samples during processing. The base MDL listed in the FLDEP SOP is based on the maximum filtration volume and minimum extract volume and will therefore be the lowest MDL. This MDL was last verified by the FLDEP laboratory 6/16/2015 (as presented in version BB-029-2.1 of the FLDEP SOP for Spectrophotometric

Determination of Corrected and Uncorrected Chlorophyll *a* and Phaeophytin, available here:
<https://fldeploc.dep.state.fl.us/sop/sop3.asp?sect=BIOLOGY&cat=CHLOROPHYLL-BOD-SEDIMENT+GRAIN+SIZE&A1=Submit>).

Base MDL values for ANERR 2015 plant pigment parameters:

Parameter	Start Date	End Date	MDL	Units	Date Revisited
CHLA_N	01/01/16	12/31/16	0.55	ug/L	6/6/2015
UncCHLA_N	01/01/16	12/31/16	0.40	ug/L	6/6/2015
PHEA	01/01/16	12/31/16	0.40	ug/L	6/6/2015

The sample MDL is calculated based on the number of times a sample must be diluted. For example, if a CHL_A sample must be diluted to twice its volume, the base MDL of 0.55 ug/L is multiplied by a dilution factor of two (0.55 ug/L x 2) thus resulting in an MDL of 1.10 ug/L. For samples that fall below the MDL and their MDL is greater than the base MDL, individual sample MDLs are listed in the table below. These data have been flagged and coded as -4 SBL in the dataset.

2016 MDLs for Chlorophyll *a* (CHLA_N), Uncorrected Chlorophyll *a* (UncCHLA_N), and Phaeophytin (PHEA), as reported by FLDEP laboratory when values differ from base MDL values:

Parameter	DateTimeStamp	Site	MDL	Units
CHLA_N	1/13/16 10:07	apawpnut	1.0	ug/L
CHLA_N	1/13/16 8:48	apambnut	0.69	ug/L
CHLA_N	1/13/16 12:43	apaegnut	1.1	ug/L
CHLA_N	1/13/16 11:41	apanhnut	1.3	ug/L
CHLA_N	1/13/16 12:13	apacpnut	1.5	ug/L
CHLA_N	1/13/16 13:18	aparvnut	1.2	ug/L
CHLA_N	1/13/16 11:44	apanhnut	1.1	ug/L
CHLA_N	1/13/16 11:48	apanhnut	1.1	ug/L
CHLA_N	4/5/16 8:15	apaesnut	1.4	ug/L
UncCHLA_N	1/13/16 10:07	apawpnut	0.75	ug/L
UncCHLA_N	1/13/16 13:18	aparvnut	0.91	ug/L
UncCHLA_N	4/5/16 8:15	apaesnut	1.0	ug/L
PHEA	1/13/16 10:07	apawpnut	0.75	ug/L
PHEA	1/13/16 9:10	apadbnut	1.0	ug/L
PHEA	1/13/16 10:48	apapcnut	0.91	ug/L
PHEA	1/13/16 8:48	apambnut	0.50	ug/L
PHEA	1/13/16 12:43	apaegnut	0.77	ug/L
PHEA	1/13/16 11:16	apascnut	0.45	ug/L
PHEA	1/13/16 11:41	apanhnut	0.92	ug/L
PHEA	1/13/16 12:13	apacpnut	1.1	ug/L
PHEA	1/13/16 13:18	aparvnut	0.91	ug/L
PHEA	1/13/16 11:44	apanhnut	0.83	ug/L
PHEA	1/13/16 11:48	apanhnut	0.82	ug/L

PHEA	1/14/16 5:00	apaesnut	0.83	ug/L
PHEA	1/14/16 10:00	apaesnut	0.67	ug/L
PHEA	2/17/16 12:36	apaegnut	0.90	ug/L
PHEA	2/17/16 10:36	apascnut	0.45	ug/L
PHEA	2/17/16 10:59	apanhnut	0.67	ug/L
PHEA	2/17/16 13:06	aparvnut	0.75	ug/L
PHEA	3/1/16 13:25	apadbnut	0.50	ug/L
PHEA	3/1/16 12:12	apapcnut	0.60	ug/L
PHEA	3/1/16 14:00	apambnut	0.90	ug/L
PHEA	3/1/16 9:24	apaegnut	0.50	ug/L
PHEA	3/1/16 11:27	apascnut	0.55	ug/L
PHEA	3/1/16 10:34	apanhnut	2.5	ug/L
PHEA	3/1/16 14:41	aparvnut	1.0	ug/L
PHEA	3/1/16 13:27	apadbnut	0.50	ug/L
PHEA	4/5/16 8:15	apaesnut	1.0	ug/L
PHEA	4/5/16 15:45	apaesnut	0.67	ug/L
PHEA	4/5/16 18:15	apaesnut	1.0	ug/L
PHEA	4/5/16 20:45	apaesnut	0.80	ug/L
PHEA	4/5/16 23:15	apaesnut	1.0	ug/L
PHEA	4/6/16 1:45	apaesnut	1.0	ug/L
PHEA	4/6/16 4:15	apaesnut	0.80	ug/L
PHEA	4/6/16 6:45	apaesnut	1.0	ug/L
PHEA	10/3/16 9:21	apawpnut	0.56	ug/L
PHEA	10/3/16 9:42	apadbnut	2.0	ug/L
PHEA	10/3/16 10:00	apambnut	0.67	ug/L
PHEA	11/1/16 9:52	apawpnut	0.90	ug/L
PHEA	11/1/16 10:19	apadbnut	0.73	ug/L
PHEA	11/1/16 9:34	apapcnut	0.73	ug/L
PHEA	11/1/16 10:41	apambnut	0.74	ug/L
PHEA	11/1/16 8:15	apaegnut	0.79	ug/L
PHEA	11/1/16 7:50	apaebnut	0.90	ug/L
PHEA	11/1/16 8:57	apanhnut	0.81	ug/L
PHEA	12/8/16 7:30	apaesnut	1.3	ug/L

13) Laboratory methods –

a) Parameter: PO4

- 1) Method Reference: The described procedure is based on EPA Method 365.1, Rev. 2.0 (1993) and the Bran+Lubbe method G-146-95 Rev. 3.

- 2) Method Description: Orthophosphate reacts with molybdenum (VI) and antimony (III) in an acid medium to form an antimony-phospho-molybdate complex. The complex is reduced with ascorbic acid to form a blue complex that absorbs at 880 nm.
- 3) Preservation Method: Samples are filtered in the field, placed on ice (not frozen), and analyzed within 48 hours of sample collection.

b) Parameter: TP

- 1) Method Reference: This SOP is based on EPA Method 365.1, Rev. 2.0 (1993) and SEAL Analytical AQ2 Method: EPA-119-A Rev. 5.
- 2) Method Description: Prior to analysis the samples are acid-persulfate digested according to the DEP SOP NU-049. This process converts inorganic and organic forms of phosphorus to ortho-phosphate. Ortho-phosphate reacts with molybdenum and antimony in an acidic medium to form a phosphoantimony/molybdenum complex, which is reduced with ascorbic acid. The AQ2 Discrete Analyzer is used to measure the absorbance of the complex at 880 nm.
- 3) Preservation Method: Samples are acidified in the field to pH <2, placed on ice (not frozen), and analyzed within 28 days of sample collection.

c) Parameter: NH4

- 1) Method Reference: This SOP is based upon EPA Method 350.1, Rev. 2.0 (1993) and OI Analytical Method 3271152 utilizing gas diffusion.
- 2) Method Description: The sample pH is raised to a pH of >11. The ammonia molecules generated at this pH pass through a gas diffusion membrane and are absorbed into an alkaline hypochlorite solution to form chloramine. The chloramine reacts with salicylate to form indophenol blue in an amount that is proportional to the ammonia concentration. Sodium nitroferrocyanide intensifies the blue color. The absorbance is measured at 660 nm.
- 3) Preservation Method: Samples are filtered in the field, acidified to pH <2, placed on ice in the dark and analyzed within 28 days.

d) Parameter: NO23

- 1) Method Reference: This method is based on EPA method 353.2, Rev 2.0 (1993) and Lachat method 10-107-04-1-C.
- 2) Method Description: A filtered sample is passed through a column containing granular copper-cadmium, which reduces nitrate to nitrite. The nitrite originally present plus the reduced nitrate can then be determined by colorimetry. The nitrite is diazotized with sulfanilamide and coupled with N-(1-naphthyl) ethylenediamine dihydrochloride to form a highly colored azo dye, which is measured at a wavelength of 520 nm.
- 3) Preservation Method: Samples are filtered in the field, acidified to pH <2, placed on ice in the dark and analyzed within 28 days.

e) Parameter: TKN

- 1) Method Reference: This SOP is based on EPA method 351.2, Rev. 2.0 (1993) and Seal AQ2 method EPA-111-A Rev. 4.
- 2) Method Description: Prior to analysis, digestion converts free ammonia and organic nitrogen compounds to ammonium sulfate (DEP SOP NU-091). Ammonium reacts with salicylate and hypochlorite in a buffered, alkaline solution in the presence of sodium nitroferrocyanide (pH = 12.4-12.7) to form the salicylic acid analog of indophenol blue. The blue-green color produced is measured at 660 nm.
- 3) Preservation Method: Whole water is acidified in the field to pH <2, placed on ice in the dark and analyzed within 28 days.

f) Parameter: CHLA_N and UncCHLA_N and PHEA

- 1) Method Reference: This method is based on Standard Methods 10200H and EPA Method 446.0.
- 2) Method Description: This method is used to determine the amount of chlorophyll-*a* and pheophytin-*a* in marine and freshwater algae by visible spectrophotometry. Uncorrected chlorophyll-*a* is calculated using the trichromatic equation. Corrected chlorophyll-*a* and pheophytin are calculated using the monochromatic equation. The

absorption-peak-ratio (chlorophyll/pheophytin) is also determined. A sample is vacuum filtered onto a glass fiber filter. The filter is then macerated with a tissue grinder and steeped in 90% acetone to extract chlorophyll from the algal cells. The sample is clarified through centrifugation. The absorbance of the clarified extract is then measured on a spectrophotometer at 750, 665, 664, 647 and 630 nm wavelengths before and after a 90 second Hydrochloric acid acidification step.

- 3) Preservation Method: Whole water is collected in brown Nalgene bottles, placed on ice in the dark, and delivered to the FLDEP lab within 36 hours for filtration.

g) Parameter: TSS

- 1) Method Reference: This method is based on Standard Methods 2540 D-1997.
- 2) Method Description: A well-mixed sample is filtered through a pre-weighed glass fiber filter. The filter and any residue are then dried to a constant weight at 103-105 °C. The filter is cooled in a desiccator, weighed and the result used to compute the TSS of the sample.
- 3) Preservation Method: Whole water is placed on ice in the dark for analysis within 7 days.

14) Field and Laboratory QAQC programs –

a) Precision

- 1) Field Variability – Field blanks (using deionized water) are included in all monthly sampling events. ANERR staff collect field triplicate samples from a successive grab sample. Triplicate samples are collected from separate grabs at one sampling station every other month, rotating through SWMP datalogger stations. There are no field triplicates collected during diel sampling.
- 2) Laboratory Variability – Method blanks and duplicate samples are run with every sample batch. Batches are groups of 20 or less samples that are analyzed concurrently. Precision is measured by Relative Percent Difference (RPD).
- 3) Inter-organizational splits – None.

b) Accuracy

- 1) Sample Spikes – At least two sample spikes are performed with each sample batch. The acceptance limits for sample or spike duplicates is a RPD of less than 20% if both results are above the PQL. Laboratory fortified blanks are run with each sample batch, acceptance limits for recovery are 85-115%.
- 2) Standard Reference Material Analysis – Check standards are included in each batch and at the beginning and end of each run. Check standard acceptance limits are 85-115% recovery. (FLDEP Central Laboratory NU-043-2.16).
- 3) Cross Calibration Exercises – None for 2016.

15) QAQC flag definitions –

QAQC flags provide documentation of the data and are applied to individual data points by insertion into the parameter's associated flag column (header preceded by an F_). QAQC flags are applied to the nutrient data during secondary QAQC to indicate data that are out of sensor range low (-4), rejected due to QAQC checks (-3), missing (-2), optional and were not collected (-1), suspect (1), and that have been corrected (5). All remaining data are flagged as having passed initial QAQC checks (0) when the data are uploaded and assimilated into the CDMO ODIS as provisional plus data. The historical data flag (4) is used to indicate data that were submitted to the CDMO prior to the initiation of secondary QAQC flags and codes (and the use of the automated primary QAQC system for WQ and MET data). This flag is only present in historical data that are exported from the CDMO ODIS.

- 4 Outside Low Sensor Range
- 3 Data Rejected due to QAQC
- 2 Missing Data

- 1 Optional SWMP Supported Parameter
- 0 Data Passed Initial QAQC Checks
- 1 Suspect Data
- 4 Historical Data: Pre-Auto QAQC
- 5 Corrected Data

16) QAQC code definitions –

QAQC codes are used in conjunction with QAQC flags to provide further documentation of the data and are also applied by insertion into the associated flag column. There are three (3) different code categories, general, sensor, and comment. General errors document general problems with the sample or sample collection, sensor errors document common sensor or parameter specific problems, and comment codes are used to further document conditions or a problem with the data. Only one general or sensor error and one comment code can be applied to a particular data point. However, a record flag column (F_Record) in the nutrient data allows multiple comment codes to be applied to the entire data record.

General errors

GCM	Calculated value could not be determined due to missing data
GCR	Calculated value could not be determined due to rejected data
GDM	Data missing or sample never collected
GQD	Data rejected due to QA/QC checks
GQS	Data suspect due to QA/QC checks
GSM	See metadata

Sensor errors

SBL	Value below minimum limit of method detection
SCB	Calculated value could not be determined due to a below MDL component
SCC	Calculation with this component resulted in a negative value
SNV	Calculated value is negative
SRD	Replicate values differ substantially
SUL	Value above upper limit of method detection

Parameter Comments

CAB	Algal bloom
CDR	Sample diluted and rerun
CHB	Sample held beyond specified holding time
CIP	Ice present in sample vicinity
CIF	Flotsam present in sample vicinity
CLE	Sample collected later/earlier than scheduled
CRE	Significant rain event
CSM	See metadata
CUS	Lab analysis from unpreserved sample

Record comments

CAB	Algal bloom
CHB	Sample held beyond specified holding time
CIP	Ice present in sample vicinity
CIF	Flotsam present in sample vicinity
CLE	Sample collected later/earlier than scheduled
CRE	Significant rain event

CSM	See metadata
CUS	Lab analysis from unpreserved sample
<i>Cloud cover</i>	
CCL	clear (0-10%)
CSP	scattered to partly cloudy (10-50%)
CPB	partly to broken (50-90%)
COC	overcast (>90%)
CFY	foggy
CHY	hazy
CCC	cloud (no percentage)
<i>Precipitation</i>	
PNP	none
PDR	drizzle
PLR	light rain
PHR	heavy rain
PSQ	squally
PFQ	frozen precipitation (sleet/snow/freezing rain)
PSR	mixed rain and snow
<i>Tide stage</i>	
TSE	ebb tide
TSF	flood tide
TSH	high tide
TSL	low tide
<i>Wave height</i>	
WH0	0 to <0.1 meters
WH1	0.1 to 0.3 meters
WH2	0.3 to 0.6 meters
WH3	0.6 to > 1.0 meters
WH4	1.0 to 1.3 meters
WH5	1.3 or greater meters
<i>Wind direction</i>	
N	from the north
NNE	from the north northeast
NE	from the northeast
ENE	from the east northeast
E	from the east
ESE	from the east southeast
SE	from the southeast
SSE	from the south southeast
S	from the south
SSW	from the south southwest
SW	from the southwest
WSW	from the west southwest
W	from the west
WNW	from the west northwest
NW	from the northwest
NNW	from the north northwest
<i>Wind speed</i>	
WS0	0 to 1 knot
WS1	> 1 to 10 knots
WS2	> 10 to 20 knots
WS3	> 20 to 30 knots

WS4 > 30 to 40 knots
WS5 > 40 knots

17) Other remarks/notes –

Data may be missing due to problems with sample collection or processing. Laboratories in the NERRS System submit data that are censored at a lower detection rate limit, called the Method Detection Limit or MDL. MDLs for specific parameters are listed in the Laboratory Methods and Detection Limits Section (Section II, Part 12) of this document. Concentrations that are less than this limit are censored with the use of a QAQC flag and code, and the reported value is the method detection limit itself rather than a measured value. For example, if the measured concentration of NO₃F was 0.0005 mg/l as N (MDL=0.0008), the reported value would be 0.0008 and would be flagged as out of sensor range low (-4) and coded SBL. In addition, if any of the components used to calculate a variable are below the MDL, the calculated variable is removed and flagged/coded -4 SCB. If a calculated value is negative, it is rejected and all measured components are marked suspect. If additional information on MDL's or missing, suspect, or rejected data is needed, contact the Research Coordinator at the Reserve submitting the data.

Note: The way below MDL values are handled in the NERRS SWMP dataset was changed in November of 2011. Previously, below MDL data from 2007-2010 were also flagged/coded, but either reported as the measured value or a blank cell. Any 2007-2011 nutrient/pigment data downloaded from the CDMO prior to November of 2011 will reflect this difference.

a) Information about flagged data and additional notes

January 2016:

- apaesnut 1/13/2016 9:00:00 (ISCO 240) sample missing total suspended solids (TSS) because bottle in ISCO automated sampler leaked, and there was not enough sample remaining to complete all analytical tests.
- apaesnut 1/14/2016 5:00:00 (ISCO 248) sample missing orthophosphate (PO₄F) because bottle in ISCO automated sampler leaked, and there was not enough sample remaining to complete all analytical tests.

February 2016:

- apaesnut 2/16/2016 09:05 CHLA_N, UncCHLA_N, and PHEA samples were delivered to the FLDEP laboratory 02/18/2016 at 09:12, which is just outside required hold times for FLDEP standards but within acceptable hold times per CDMO standards. Data were not flagged as suspect.
- apaesnut 2/16/2016 11:35 (ISCO 241) orthophosphate (PO₄F) and total phosphorus (TP) flagged suspect <1> during QAQC checks. Parameter levels elevated above all other diel samples for this month's diel sampling and are high values for the site in general, and other parameters do not reflect a change in water conditions leading us to suspect contamination.

April 2016:

- apaesnut 4/5/2016 8:15:00 (ISCO 240) sample missing total suspended solids (TSS) because there was not enough sample remaining to complete all analytical tests.
- apaesnut 4/6/2016 9:15:00 (ISCO 250) sample not taken because automated diel sampler was retrieved prior to final sample collection. It is flagged and coded as <-2> GDM.

May 2016:

- apaebnut 5/10/2016 7:56:00 (191 East Bay Bottom) Nitrate + Nitrite NO₂3F sample flagged as suspect because of faulty filter apparatus.

- Apawpnut 5/10/2016 10:06:00 (131 West Pass) orthophosphate (PO₄F) flagged suspect <1> during QAQC checks. Orthophosphate levels elevated above typical levels for the site and other parameters do not reflect a change in water conditions.
- apascnut (Sikes Cut 201) grab sample not collected due to poor weather conditions.

July 2016:

- apaesnut 7/6/2016 09:40 (ISCO 250) sample not taken because automated diel sampler was retrieved prior to final sample collection. It is flagged and coded as <-2> GDM.

September 2016:

- Poor weather conditions during grab sample collections prevented all station from being sampled 9/12/2016. Additional grab samples from some remaining stations were collected 9/13/2016, but bad weather conditions prevented collection of samples from the apawpnut (West Pass 131) and apascnut (Sikes Cut 201) for the month of September.

October 2016:

- apascnut 10/03/2016 Ammonium (NH₄F) flagged suspect <1> during QAQC checks. Ammonium levels elevated above typical levels for the site and other parameters do not reflect a change in water conditions.

November 2016:

- Automated ISCO sampler malfunctioned on 11/1/2016. It was redeployed 11/7/2016, which is why grab and diel samples were collected asynchronously for the month of November. apascnut (Sikes Cut 201) grab sample not collected due to poor weather conditions.
- apadbnut 11/1/2016 10:19:00 (141 Dry Bar) Total Kjeldahl Nitrogen (TKN) and Total Phosphorus (TP) flagged as suspect due to improper sample preservation.

December 2016:

- apaesnut 12/8/2016 0:00 and 05:00 (ISCO 246 and 248) orthophosphate (PO₄F) flagged suspect <1> during QAQC checks. Orthophosphate levels elevated above all other diel samples for this month's diel sampling and the rest of the year at this station, and other parameters do not reflect a change in water conditions leading us to suspect contamination.

b) Sample hold times

Sample Descriptor	Date Analyzed						
	NH4F	NO23F	PO4F	CHLA_N, UncCHLA_N, PHEA	TKN	TP	TSS
1/13/2016, all grab samples	1/26/2016	1/21/2016	1/14/2016	1/21/2016	1/21/2016	1/22/2016	1/19/2016
1/16/-1/14/2016, all diel samples	1/26-28/2016	1/27/2016	1/14/2016	1/22/2016	1/25/2016	1/27/2016	1/19/2016
2/17/2016, all grab samples	2/25/2016	2/23/2016	2/18/2016	3/1/2016	2/24/2016	2/24/2016	2/19/2017
2/16-17/2016, all diel samples	2/25/2016	2/26/2016	2/18/2016	3/2/2016	2/26/2016	2/24/2016	2/19/2016
3/1/2016, all grab samples	3/10/2016	3/8/2016	3/2/2016	3/15/2016	3/14/2016	3/14/2016	3/7/2016
3/1-2/2016, all diel samples	3/15/2016	3/8/2016	3/2/2016	3/15/2016	3/14/2016	3/14/2016	3/7/2016
4/5/2016, all grab samples	4/13/2016	4/18/2016	4/6/2016	4/16/2016	4/19/2016	4/15/2016	4/11/2016
4/5-6/2016, all diel samples	4/13/2016	4/16/2016	4/6/2016	4/16/2016	4/21/2016	4/15/2016	4/11/2016
5/10/2016, all grab samples	5/24/2016	5/17/2016	5/11/2016	5/17/2016	5/23/2016	5/25/2016	5/13/2016
5/10-11/2016, all diel samples	5/18/2016	5/23/2016	5/11/2016	5/17/2016	5/25/2016	5/27/2016	5/13/2016
6/1/2016, all grab samples	6/9/2016	6/6/2016	6/2/2016	6/15/2016	6/8/2016	6/14/2016	6/6/2016
6/1-2/2016, all diel samples	6/9/2016	6/6/2016	6/2/2016	6/15/2016	6/9/2016	6/14/2016	6/6/2016
7/5/2016, all grab samples	7/11/2016	7/8/2016	7/6/2016	7/18/2016	7/11/2016	7/12/2016	7/8/2016
7/5-76/2016, all diel samples	7/11/2016	7/11-12/2016	7/6/2016	7/19/2016	7/12/2016	7/12/2016	7/8/2016
8/2/2016, all grab samples	8/4/2016	8/12/2016	8/3/2016	8/18/2016	8/15/2016	8/15/2016	8/5/2016
8/2-3/2016, all diel samples	8/4/2016	8/22/2016	8/3/2016	8/18/2016	8/15/2016	8/15/2016	8/5/2016
9/12-13/2016, all grab samples	9/14/2016	9/23/2016	9/13/2016	9/23/2016	9/19/2016	9/23/2016	9/15/2016
9/12-13/2016, all diel samples	9/14/2016	9/23/2016	9/13/2016	9/23/2016	9/20/2016	9/23/2016	9/15/2016
10/3/2016, all grab samples	10/13/2016	10/7/2016	10/4/2016	10/19/2016	10/17/2016	10/18/2016	10/8/2016
10/3-4/2016, all diel samples	10/13/2016	10/12/2016	10/4/2016	10/19/2016	10/17/2016	10/18/2016	10/8/2016
11/1/2016, all grab samples	11/10/2016	11/4/2016	11/2/2016	11/14/2016	11/18/2016	11/15/2016	11/4/2016
11/7-8/2016, all diel samples	11/15/2016	11/14/2016	11/8/2016	11/16/2016	11/22/2016	11/18/2016	11/11/2016
12/7/2016, all grab samples	12/14/2016	12/13/2016	12/8/2016	12/14/2016	12/15/2016	12/14/2016	12/13/2016
12/7-8/2016, all diel samples	12/13/2016	12/13/2016	12/8/2016	12/13/2016	12/15/2016	12/14/2016	12/13/2016

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